

**Nucleic Acid Extraction Kit HandBook** 



VIRAL NUCLEIC ACID EXTRACTION USER GUIDE (Version 5.1)



FOR DIAGNOSTICS USE & RESEARCH USE ONLY

Catalog No.

SAMPLE: 5 preps GF-RD-025: 25 preps GF-RD-050: 50 preps GF-RD-100: 100 preps

GF-RD-300: 300 preps

High Yield and Purity
Fast and Easy purification
Reliable and Reproducible
in downstream applications

Eluted nucleic acid ready for use in downstream applications

No toxic or organic-based extraction required

# GF-1 Viral Nucleic Acid Extraction Kit

For isolation of viral nucleic acid (DNA/RNA) from biological samples





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## Introduction

The **GF-1 Viral Nucleic Acid Extraction Kit** is designed for rapid and efficient purification of viral DNA/RNA from samples such as serum, plasma, body fluid or virus-infected cell culture supernatant. The purification is based on the usage of denaturing agents to provide efficient viral lysis, denaturation of proteins and subsequent release of DNA or RNA. Special buffers provided in the kit are optimized to enhance the binding of DNA or RNA onto a specially-treated glass filter membrane for efficient recovery of highly pure DNA or RNA.

#### Intended Use

The **GF-1 Viral Nucleic Acid Extraction Kit** is designed for isolation and purification of viral DNA/RNA from biological samples such as serum, plasma, body fluid, saliva, biological swabs or biological cells in VTM and virus-infected cell culture supernatant. For research and in vitro diagnostic purpose only.

## **Principle of Test**

The **GF-1 Viral Nucleic Acid Extraction Kit** consists of different special buffers and GF-1 columns with silica-based membrane. The simple 4-step procedure that involves lysis, binding, washing and elution is designed for efficient isolation of nucleic acid (DNA/RNA) from a broad range of viruses. Nevertheless, different virus species will have different yield performance. Procedures must be optimized by the user.

# **Quality Control**

Each lot of **GF-1 Viral Nucleic Acid Extraction Kit** has been tested against predetermined specifications to ensure consistent product quality under ISO13485:2016 and ISO9001:2015 – certified Quality Management System.

# Kit components

Product Catalog No.	5 Preps SAMPLE	25 Preps GF-RD-025	50 Preps GF-RD-050	100 Preps GF-RD-100
Components				
GF-1 columns	5	25	50	100
Collection tubes	5	25	50	100
Buffer VL	1.5ml	6ml	12ml	24ml
Wash Buffer 1 (concentrate)*	1.5ml	7ml	14ml	28ml
Wash Buffer 2 (concentrate)*	1.7ml	9ml	17ml	36ml
Carrier RNA*	0.3mg	0.5mg	1mg	2 x 1mg
Elution Buffer	1.5ml	2 x 1.5ml	8ml	20ml
Proteinase K*	0.26ml	1.3ml	2 x 1.3ml	3 x 1.7ml
Handbook	1	1	1	1

<sup>\*</sup> Please refer to Reconstitution of Solutions and Storage and Stability.

The **GF-1 Viral Nucleic Acid Extraction Kit** is available as 25, 50 and 100 purifications per kit.

The reagents and materials provided with the kit are for diagnostics use and research purposes only.

# **Sample Material**

Tested sample types:

- 1. Blood, serum, plasma (fresh or frozen biological specimen)
- 2. Body fluid, saliva
- 3. Biological swabs or biological cells in VTM
- 4. Virus-infected cell culture supernatant

# **Storage and Stability**

- Store all solution at 20°C-30°C.
   Store Wash Buffer at room temperature with bottle capped tight after use.
- Proteinase K and Carrier RNA are stable for up to 1 year after delivery when stored at room temperature or 4°C.
   To prolong the lifetime of Proteinase K and carrier RNA, storage at -20°C is recommended.
  - To prolong the lifetime of Proteinase K and carrier RNA, storage at -20°C is recommended. **Carrier RNA solution** (after being reconstituted) can only be thawed not more than once.
- 3. Kit components are guaranteed to be stable for 18 months from the date of manufacture.
- 4. Buffer VL and Wash Buffer 1 may exhibit salt precipitation due to cold temperature. If this occurs, simply warm the bottle at 55°C 65°C with occasional mixing until completely dissolved.
- 5. Any remaining **Buffer VL** which contains **Carrier RNA** can only be stored at 4°C for not more than one week.

#### **Chemical Hazard**

**Buffer VL** and **Wash Buffer 1** contain guanidine salts which can be harmful when in contact with skin or swallowed. Always wear gloves and practice standard safety precautions. Do NOT disinfect guanidine or extraction waste in solutions containing bleach or any form of acid. To clean any items contaminated with the reagent, simply soak in detergent and water to remove all traces of guanidine before cleaning with bleach or acidic solutions.

#### Limitation

The **GF-1 Viral Nucleic Acid Extraction Kit** is intended for *in vitro* diagnostic purpose and research use only. The kit is not for the detection, prevention or treatment of a disease. Advise to do spectrophotometric and gel analysis on the extracted nucleic acid for downstream application.

#### Reconstitution of Solutions

The bottle labeled **Wash Buffer 1** and **Wash Buffer 2** contain concentrated buffer which must be diluted with absolute ethanol (>95%) before use.

# For SAMPLE (5 preps),

Add 1.5ml of absolute ethanol into the bottle labeled Wash Buffer 1.

Add 4ml of absolute ethanol into the bottle labeled Wash Buffer 2.

Add **0.3ml** of **Elution Buffer** into the vial of **Carrier RNA** and mix well, prepare inaliquots to avoid repeated freeze-thaw cycles. Store at -20°C.

# For **GF-RD-25 (25 preps)**,

Add 7ml of absolute ethanol into the bottle labeled Wash Buffer 1.

Add 21ml of absolute ethanol into the bottle labeled Wash Buffer 2.

Add **0.5ml** of **Elution Buffer** into one of the vials of **Carrier RNA** and mix well, prepare in 15µl aliquots to avoid repeated freeze-thaw cycles. Store at -20°C.

# For **GF-RD-50** (50 preps),

Add 14ml of absolute ethanol into the bottle labeled Wash Buffer 1.

Add 40ml of absolute ethanol into the bottle labeled Wash Buffer 2.

Add **1ml** of **Elution Buffer** into one of the vials of **Carrier RNA** and mix well, prepare in 15µl alignots to avoid repeated freeze-thaw cycles. Store at -20°C.

# For GF-RD-100 (100 preps),

Add 28ml of absolute ethanol into the bottle labeled Wash Buffer 1.

Add 84ml of absolute ethanol into the bottle labeled Wash Buffer 2.

Add **1ml** of **Elution Buffer** into one of the vials of **Carrier RNA** and mix well, prepare in  $15\mu l$  aliquots to avoid repeated freeze-thaw cycles. Store at -20°C. Store the other vial of **Carrier RNA** at -20°C and dissolve in **Elution Buffer** only prior to use.

#### **Procedures**

#### Reminder

- · All steps are to be carried out at room temperature unless stated otherwise.
- \* Wash Buffer 1 and Wash Buffer 2 (concentrate) have to be diluted with absolute ethanol before use. Please refer to Reconstitution of Solutions.
- If precipitation forms in **Buffer VL**, incubate at 55°C 65°C with occasional mixing until completely dissolved.
- · Pre-set waterbath to 65°C.
- Prepare Buffer VL with Carrier RNA by adding 15µl of Carrier RNA into 200µl of Buffer VL per sample.

#### 1. Sample lysis

Add 50µl of **Proteinase K** into 200µl of sample and mix thoroughly. Add 215µl of **Buffer VL** (containing **Carrier RNA**) and mix homogeneously by pulsed-vortexing. Incubate at 65°C for 10 min

#### 2. Addition of ethanol

Add 280µl of absolute ethanol. Mix immediately and thoroughly.

Mix immediately to prevent any uneven precipitation of nucleic acid due to high local ethanol concentrations.

# 3. Loading to column

Transfer the sample into a column assembled in the collection tube (provided). Centrifuge at 5,000 x g for 1 min. Discard flow through.

# 4. Column washing 1

Wash the column with  $500\mu$ l **Wash Buffer 1** and centrifuge at  $5,000 \times g$  for 1 min. Discard flow through.

Ensure that ethanol has been added into the Wash Buffer 1 before use (refer to Reconstitution of Solutions).

# 5. Column washing 2

Wash the column with 500µl **Wash Buffer 2** and centrifuge at 5,000 x g for 1 min. Discard flow through. Wash column again with 500µl **Wash Buffer 2** and centrifuge at maximum speed for 3 min.

Ensure that ethanol has been added into the Wash Buffer 2 before use (refer to Reconstitution of Solutions). Perform centrifugation for 3 min to remove ethanol.completely.

#### 6. DNA Elution

Place the column into a clean microcentrifuge tube. Add 30-50µl of **Elution Buffer** or nuclease- free water directly onto column membrane and stand for 2 min. Centrifuge at 5,000 x g for 1 min to elute DNA/RNA.

Ensure that the Elution Buffer is dispensed directly onto the center of the membrane for complete elution. Store DNA at 4°C to -20°C or RNA at -20°C to -80°C.

# **Troubleshooting**

Please note that by not adhering to the recommended protocols, unsatisfactory results related to yield and quality of DNA/RNA may occur. If problems arise, please refer to the following:

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Problem	Possibility	Suggestions
Low DNA/RNA yield	Samples not fresh or not properly stored	Sample can only be thawed not more than once.
	Carrier RNA is not added to Buffer VL	Prepare Buffer VL with Carrier RNA as described in the procedures page.
	Low quality of Carrier RNA	Ensure that the Carrier RNA is aliquoted and can only be thawed not more than once. Please refer to page 3 for the "Storage and Stability".
		Ensure that any precipitate formed in <b>Buffer VL</b> is completely dissolved.
	Inefficient nuclease inhibition during sample lysis step	Ensure that <b>Buffer VL</b> is mixed homogeneously with the mixture of sample and <b>Proteinase K</b> .
	Ethanol is not added after sample lysis	Repeat purification with new sample.
	Wash Buffer 1 and Wash Buffer 2 are reconstituted wrongly	Please refer to 'Reconstitution of Solutions". Repeat purification with new sample.

# Troubleshooting

Problem	Possibility	Suggestions
	Column is not dried before addition of Elution Buffer	Ensure that column is spun dried maximum speed for 3 minutes after addition of Wash Buffer 2.
Poor performance of eluted DNA/RNA in downstream applications	RNA degraded	Process sample immediately or sample is stored for later use, ensure that sample is thawed on ice.
		Use disposable plasticware and pipette tips.
		Ensure that the purification is performed in an RNase-free environment.
	Eluted DNA/RNA contains traces of ethanol	Ensure that the Column drying step is carried out prior to elution.
	Low concentration of eluted DNA/RNA	Reduce the amount of Elution Buffer but not less than 30µl
	The amount of added carrier RNA is inappropriate	User may optimize the amount of Carrier RNA to be added.

# Addition of ethanol Sample Lysis Add 50µl Proteinase K into 200µl of Add 280ul absolute sample and mix thoroughly. Add 215µl of ethanol and mix Buffer VL (containing Carrier RNA) and immediately. mix by pulsed vortexing. Incubate at 65°C,10 min $5.000 \times g$ 1 min Column Washing Loading to column Centrifuge Centrifuge Transfer sample Add 500µl Wash Buffer 1 Discard flow through Discard flow through to column 3 min 1 min Column Washing Centrifuge Column Washing Centrifuge Add 500 µl Wash Buffer 2 Discard flow through Add 500µl Wash Buffer 2 Discard flow through **Elution**



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#### Manufacturing

Transfer column to a new

microcentrifuge tube

Add 30-50ul Elution Buffer or water. Stand for 2 min.

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Centrifuge

Store DNA at 4°C or -20°C

Store RNA at -20°C or -80°C

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